

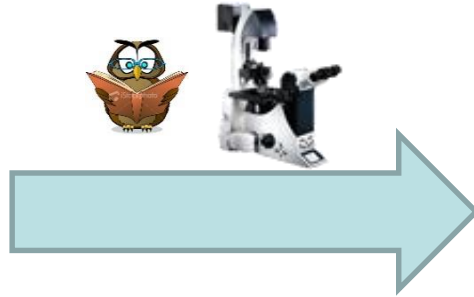


CMB 551 Module 1A

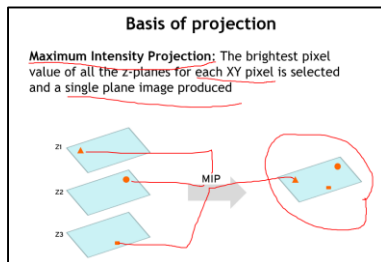
Image processing and quantitative image analysis

Sam Johnson

Benjamin Carlson

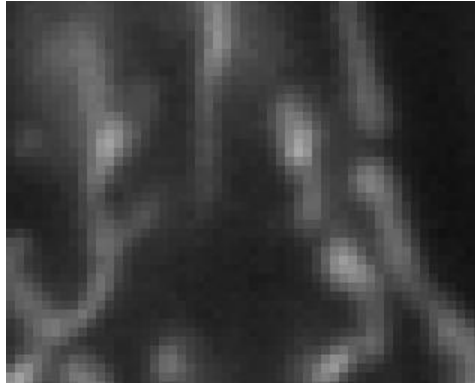


n



File:
Filename_to_open

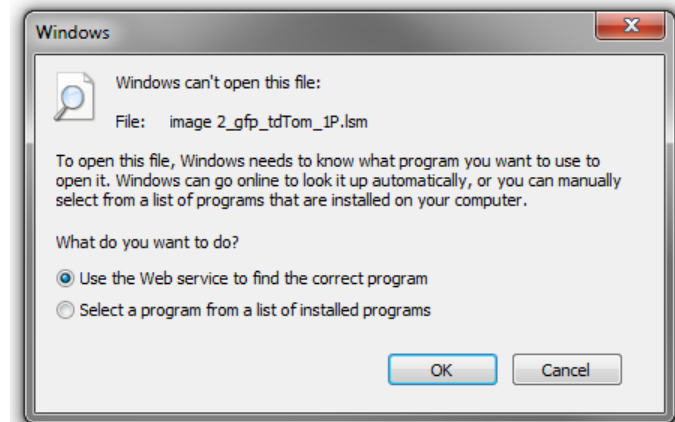
Microscopy images are somewhat different to other fields

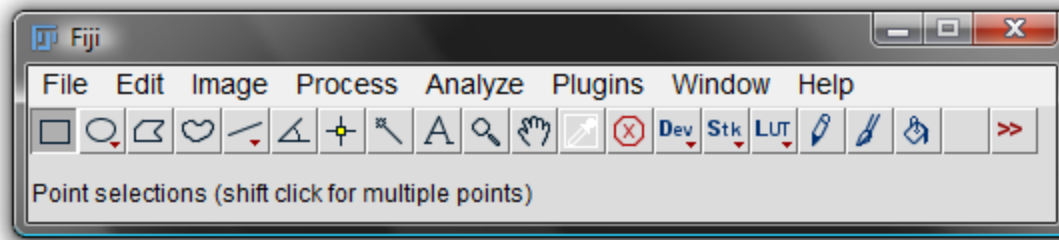


Microscopy data are in many formats

Microscope specific formats

LIF, LEI, LSM, ZVI, STK, OIB, OIF, dv, ICS/IDS, ICS2, r3d, PIC, IPL, CXD, SDT, IPW, ND2, RAW, IMS. . .





- ✓ Types of images - bitmap/vector and compression
- ✓ Bit-depth, histogram, scaling and gamma
- ✓ Export to standard formats for figures
- ✓ Color use and misuse
- ✓ 3D images - Projections and other views
- ✓ Stitching and alignment
- ✓ Image processing - Filters
- ✓ Segmentation - Regions of interest
- ✓ Quantification - Count, measure, intensity, colocalization, 3D measurements, tracking

Data → big(ish) data

One image	= ~1 MB
$\lambda * 3 \quad Z * 11 \quad t * 360$	= ~12 GB

Can be a challenge to

- **Move** the data
- **Store** all your original data
- Keep it **safe**
- Be able to **find** it again

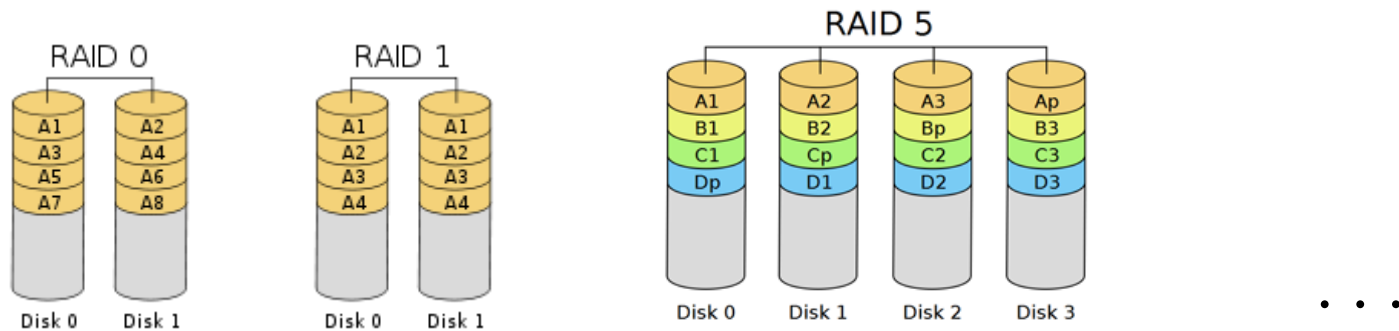


Some useful things for the microscopist to know about hard-drives and servers

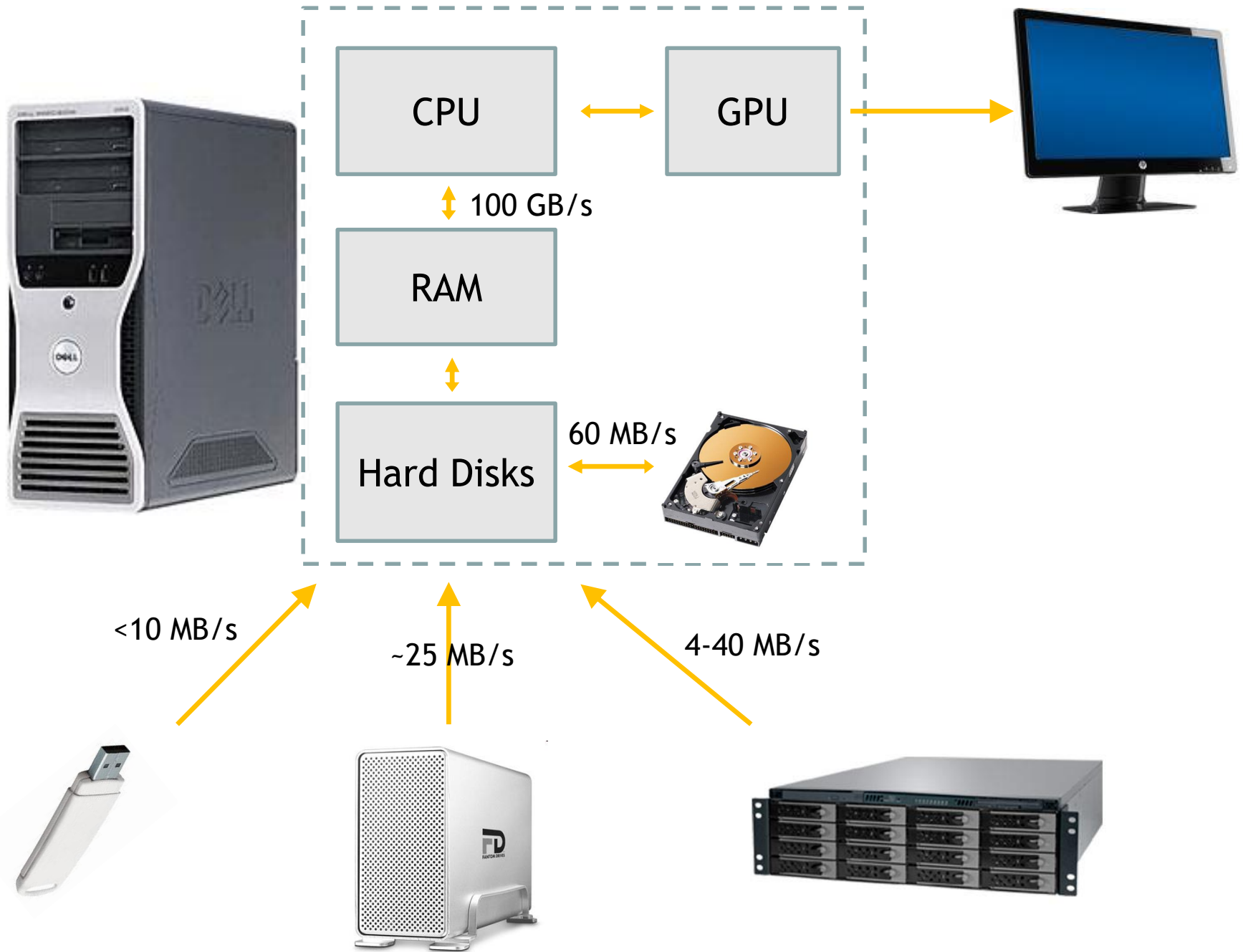
Drive formats:

- NTFS = PC (and Linux understands)
- HFS+ = Mac OS X Extended
- Ext* = Linux
- FAT32 = All (but with some limitations - 4GB max file size)

Hard-drives fail, RAID helps (not always)



Non-redundant, faster write speeds



Programs for viewing your microscope images on your computer



Free basic versions of the confocal software for the particular format



(but FIJI opens them all)



Irfanview opens 16-bit TIFF (PC only)

Are we going to use Photoshop?

No

Pho·to·shop [foh-toh-shop]

verb (used with object), Pho-to-shopped, Pho-to-shop-ping.

(sometimes lowercase) to alter (an image) using this software: *Her face is nicely Photoshopped in the ad. They've photoshopped the car onto an image of a beautiful beach.*



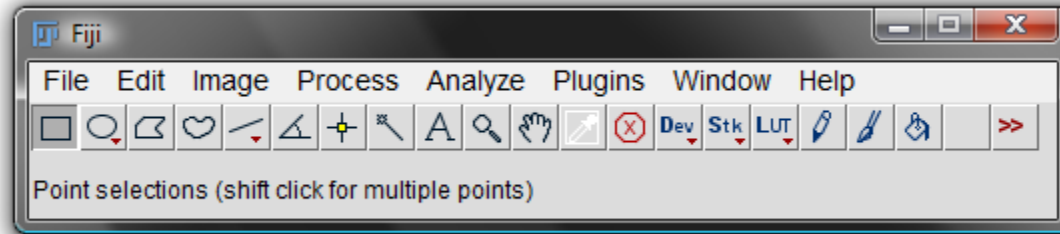
GNU Image Manipulation Program

<http://www.gimp.org/>

FIJI/ImageJ and reasons to use it

FIJI - FIJI Is Just ImageJ

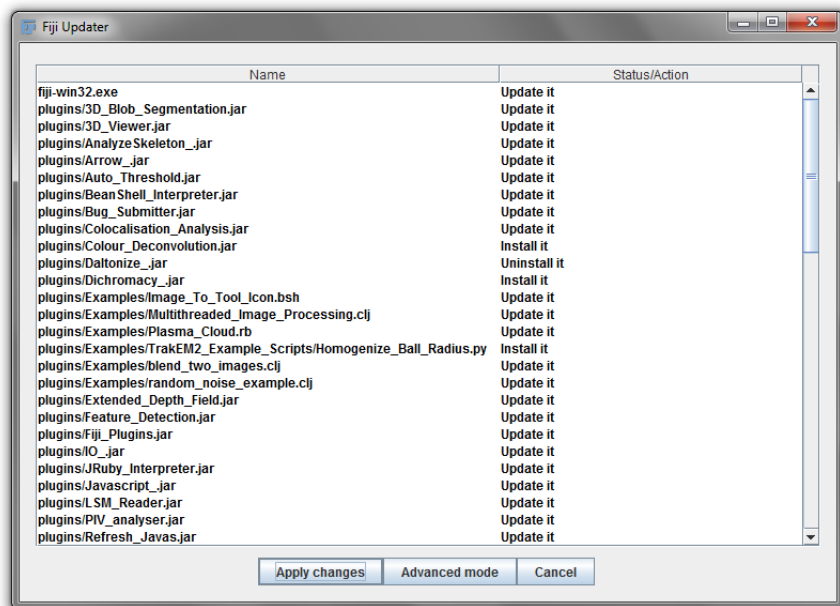
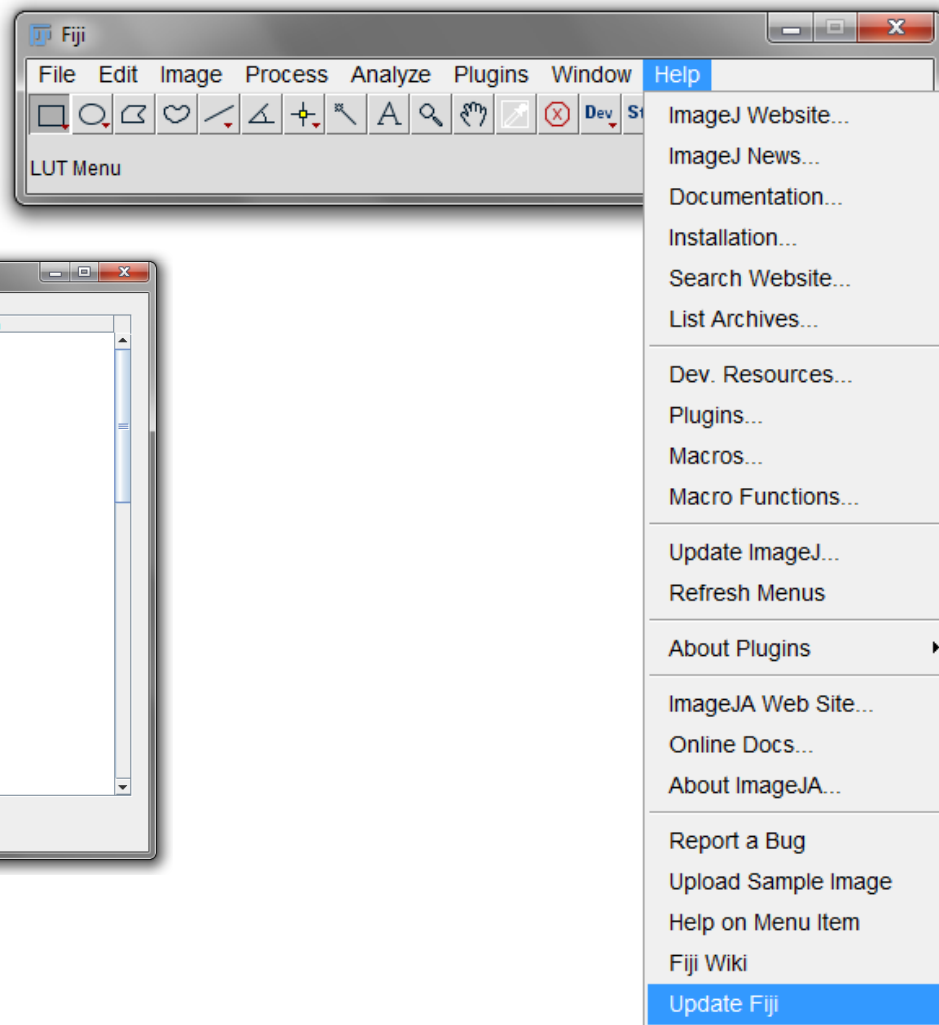
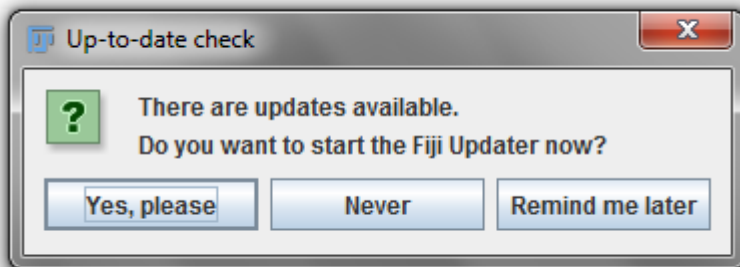
<http://fiji.sc/wiki/index.php/Fiji>



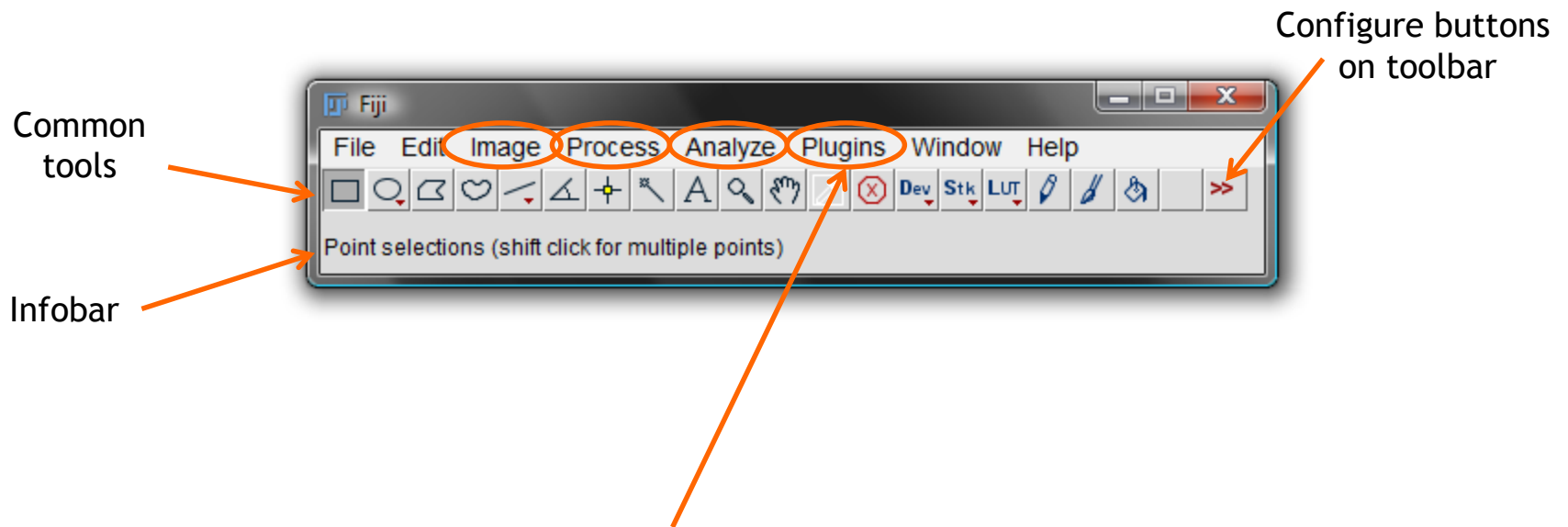
(FIJI is ImageJ + a set of useful plugins, an update tool, 3D Java, scripting and . . .)

- It's free (no cost, no licenses to deal with)
- Opens nearly anything
- It is extremely useful - many and powerful features
- It works on any OS
- Open source - (can view and modify the source code - you know what it is doing)
- Active development - lots of other people like you using it
- Has an incredible number and variety of plugins





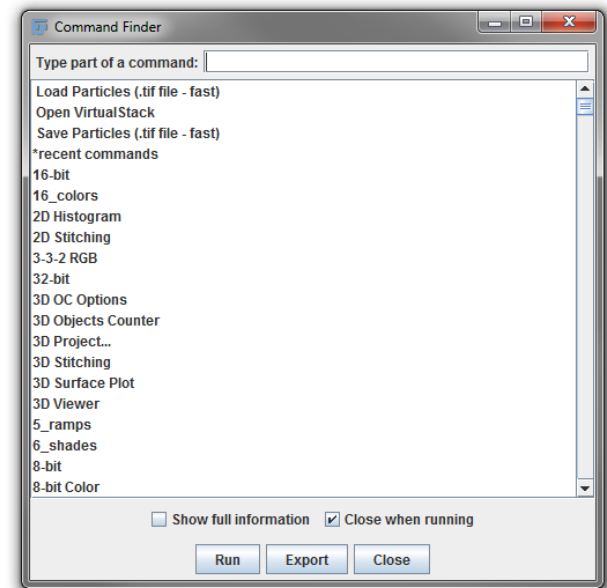
Overview of the FIJI software



Plugins and Macros are extensions of core functions - write them yourself or use ones written by other people

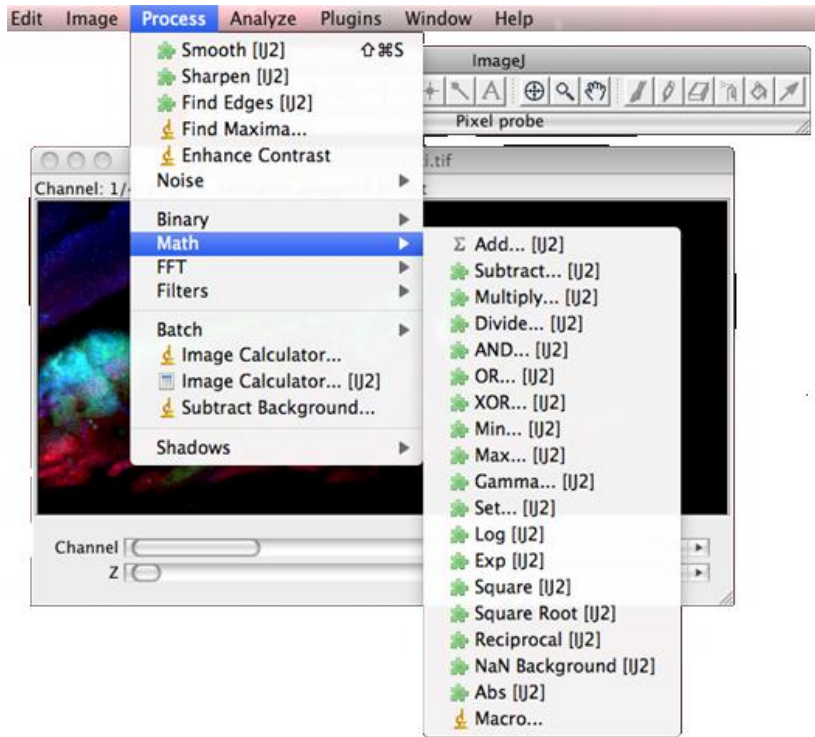
Oddities of the FIJI software

- Multiple Document Interface - lots of little windows
File/Close All is quite useful
- Java based - System and Java clipboards
- Undo - doesn't always work - *File/Revert (ctrl-R)*
- *Ctrl-L* gives command Finder

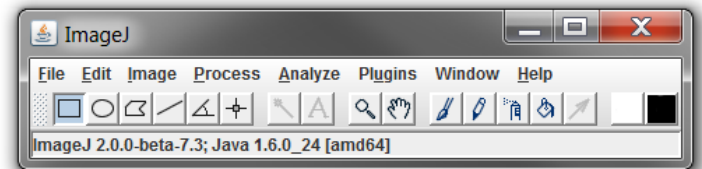


ImageJ2

<http://developer.imagej.net/downloads>



Help/Switch To Modern Mode

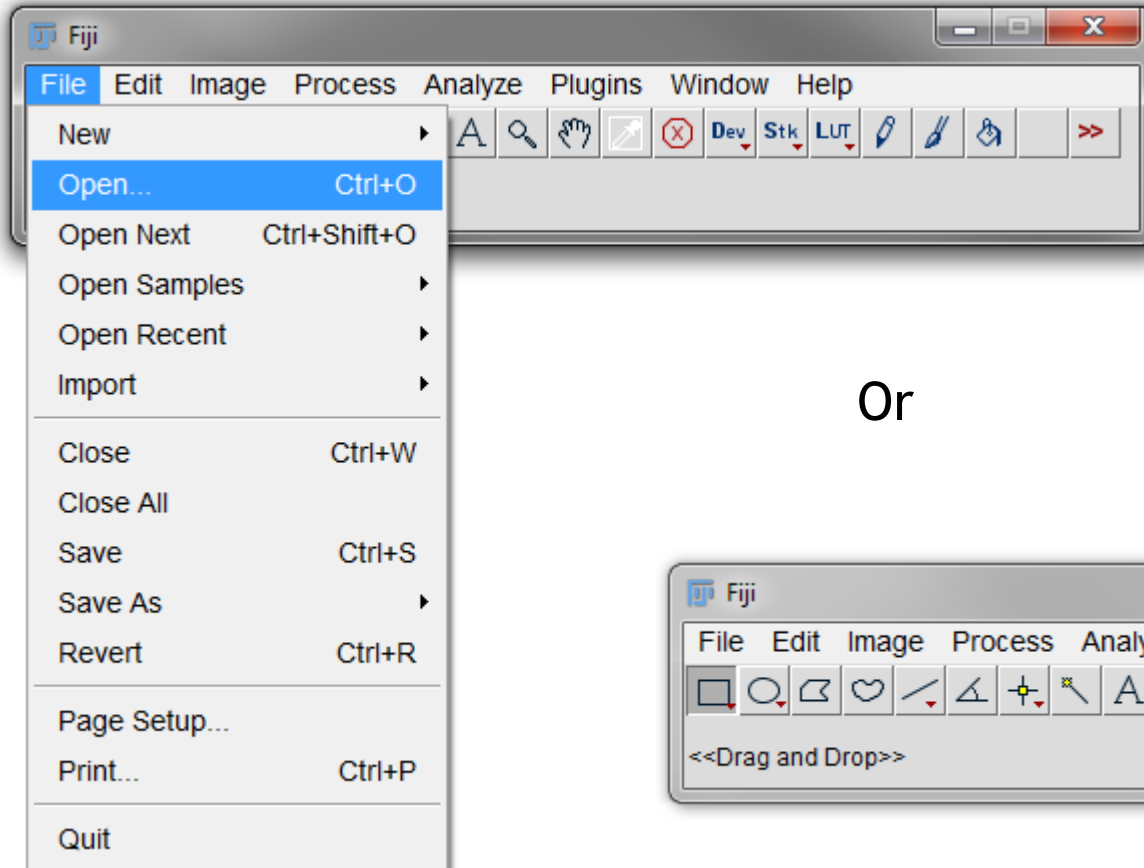


Headless | N-dimensional formats | Complete re-write

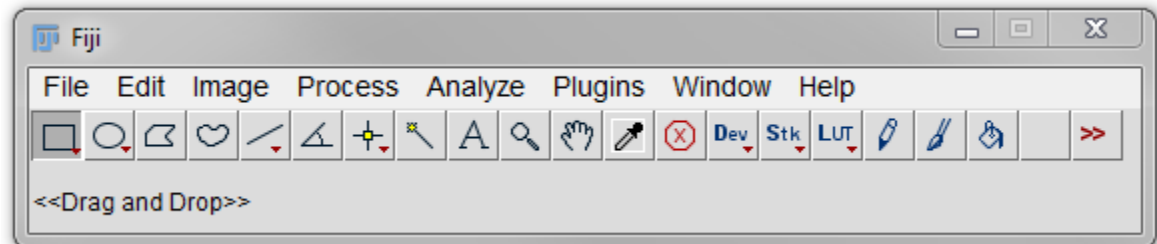


Opening a file

File:
FirstImage.tif



Or





Really basic functions

+ = Zoom in

- = Zoom out

5 = 1:1 zoom

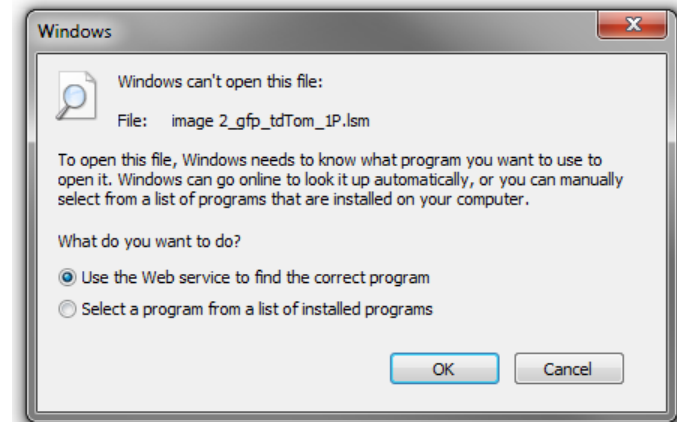
Hold space bar and left click drag = Pan

Duplicate the image:
/Image/Duplicate (or ctrl+shift+D on a PC)

Images are in many formats

Microscope specific formats

LIF, LEI, LSM, ZVI, STK, OIB, OIF, dv, ICS/IDS, ICS2, r3d, PIC, IPL, CXD, SDT, IPW, ND2, RAW, IMS. . .



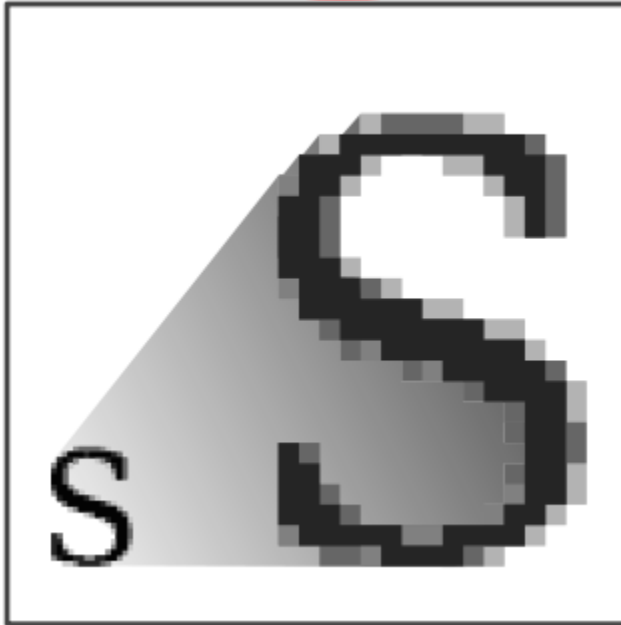
General formats

TIF, GIF, BMP, PNG, SVG, JPEG, MPEG, QuickTime, AVI, SWF, PICT, EPS, PDF, PSD, XCF . . .

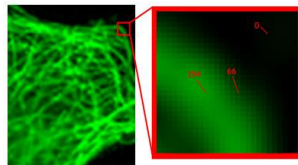
Which of these are good and what characteristics do they have?

Two types of images

Bitmap/raster



Array of pixels



Photoshop/GIMP

Vector



Mathematical/geometric equations

Scalable






Efficient for some things

Illustrator/Inkscape



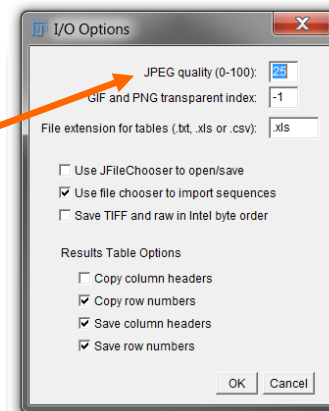
Compression

Files:
These ones

Name	Size
 File_TIF	679 KB
 File_PNG	434 KB
 File_TIF	425 KB
 File_JPEG100	142 KB
 File_JPEG25	10 KB

- How different do the images look?
- Which ones are wise to use?

(I made the JPEGs different using
Edit/Options/InputOutput



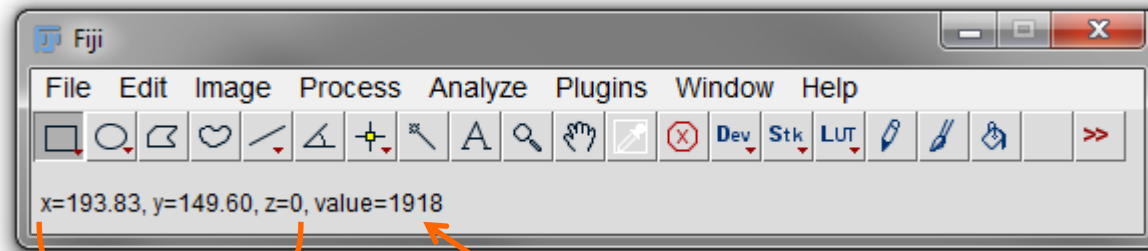
Compression types

Lossless	Lossy
Encodes exactly the original data (encodes redundant information and some other clever tricks)	Throws some information away to reduce the file size
You can get back the original	You <u>can not</u> get the original back
Sometimes not that much smaller	Allows a much smaller file, Range of quality/size
Zip etc (takes time), LZW	
PNG	JPEG

**Intensity range and
displaying them
optimally and fairly**

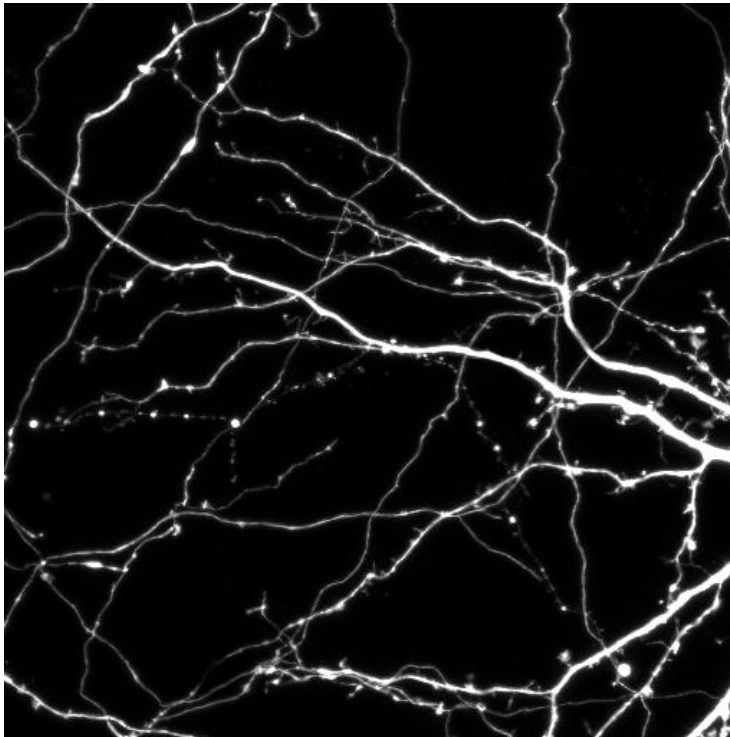
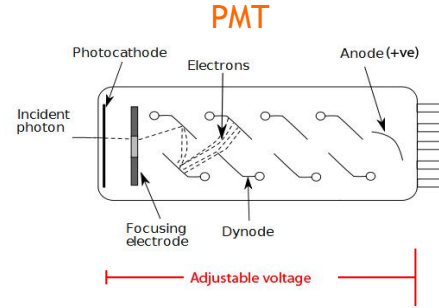
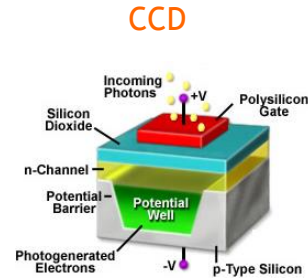


What intensities are in an image?

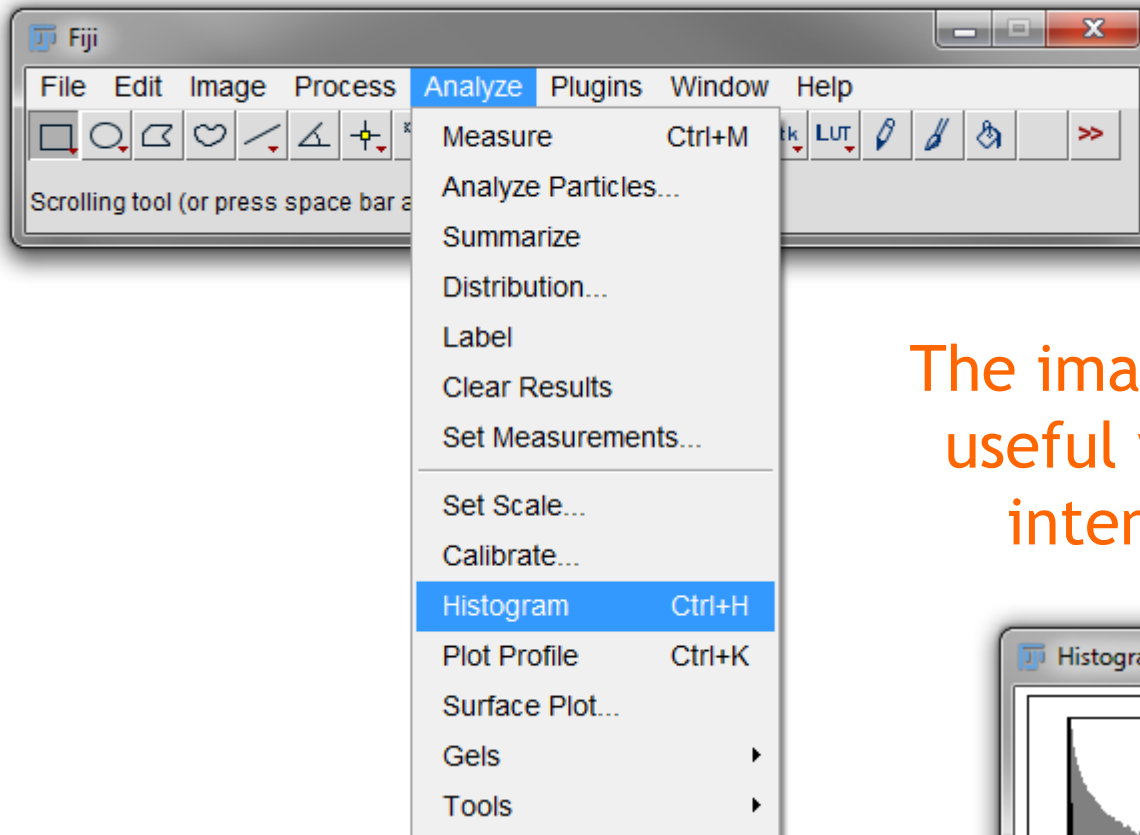


Mouse over for: Coordinates | Intensity

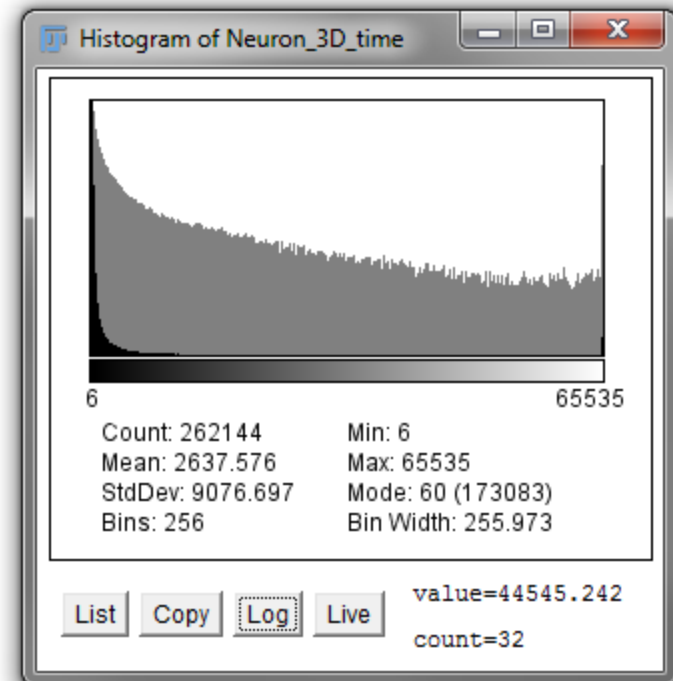
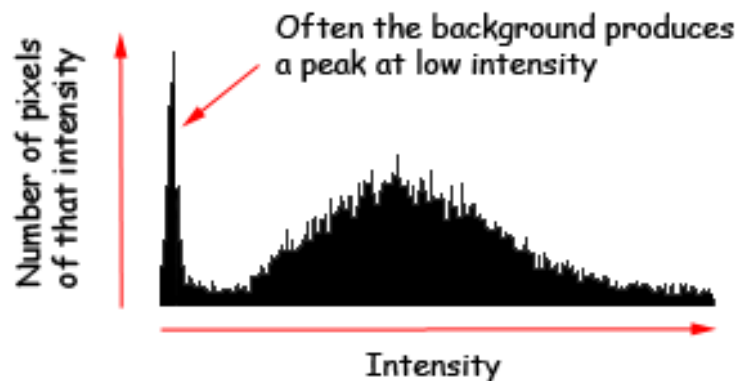
Analog to digital conversion



Histogram

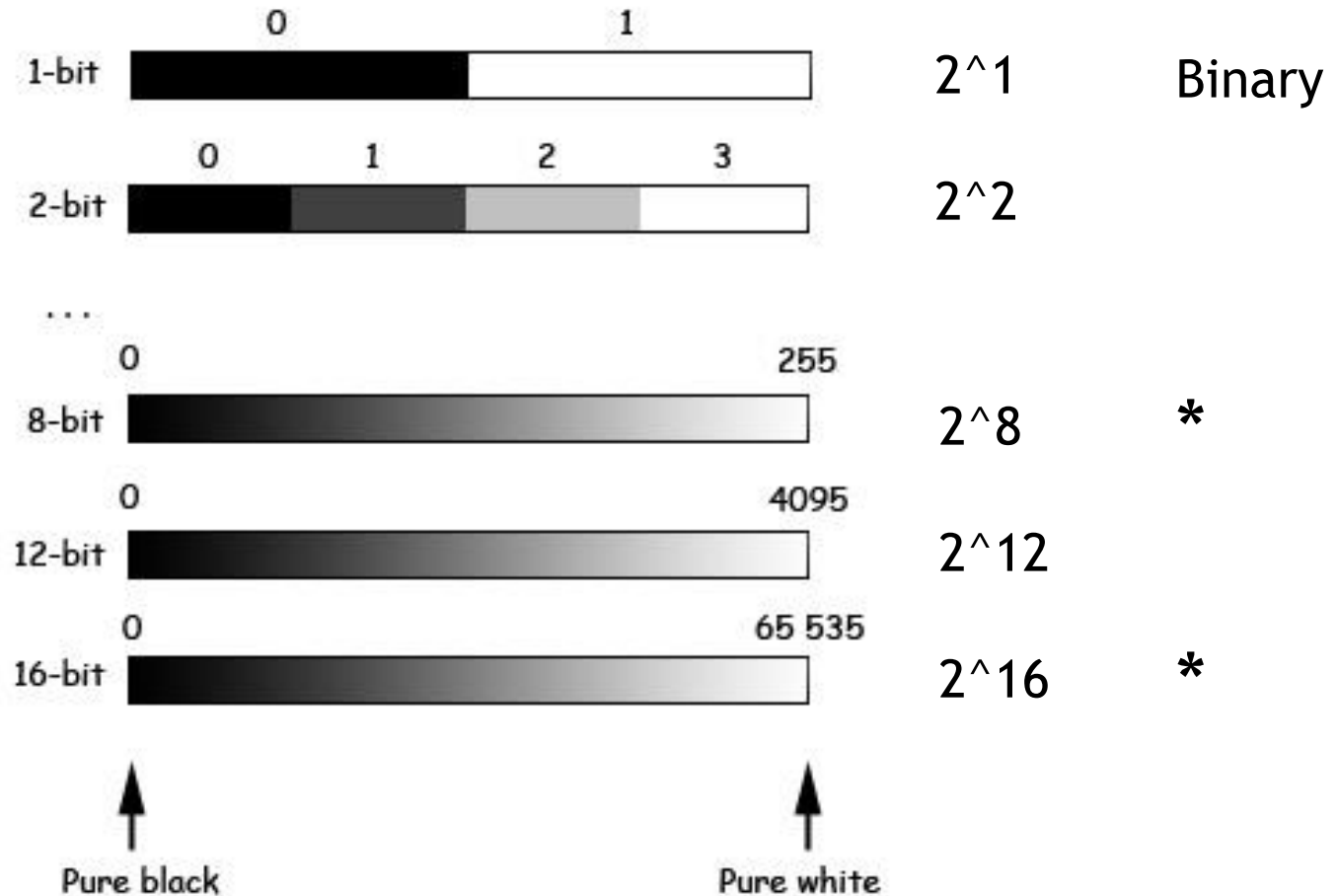


The image histogram is a very useful way of looking at the intensities in the image



Bit-depth

The range of intensities can be represented with different numbers of gray-levels



Even more Bit-depths

- Signed and unsigned 16 bit
 - 0-65,535
 - -32,768 to + 32,767



(What is a negative intensity?)

- 32-bit float - 4.3 billion + Decimals + NotaNuNumber

How many grey levels do we need?

8-bit enough for most things:

Human eye (without adaptation)

Monitors - contrast factor

Printers

But it is useful to
use more in
microscopy?

**Dynamic
range**

What span of a measurement can be measured?

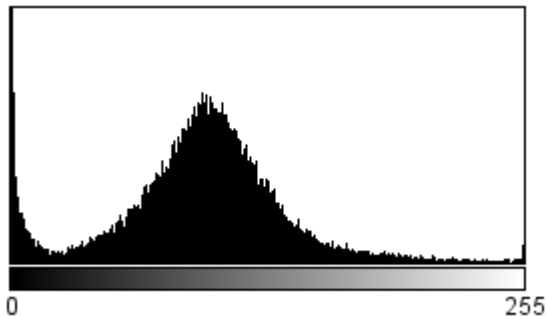
Signal at the greatest intensity of light measurable
Lowest intensity (Noise limited)

How many grey levels do we need?

How many photons/px are involved?

Say, <100

Confocal



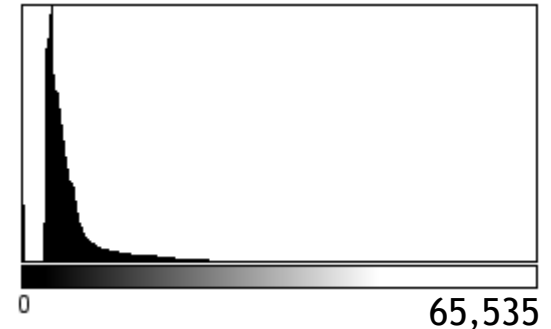
~1000

CCD



10-100 before EM

EM-CCD

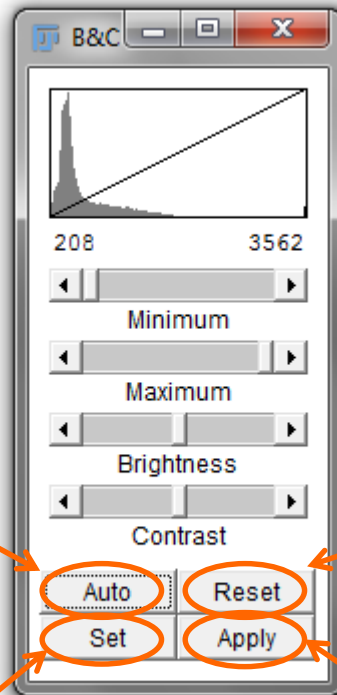
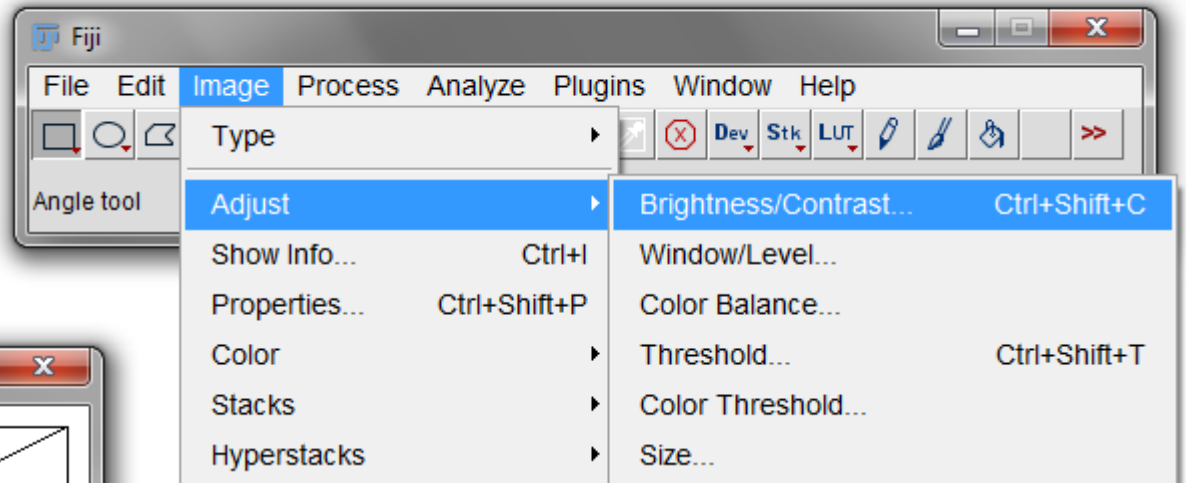


What if only some of the range is filled?



Scaling

File:
B_Imageforscaling



Enhance
contrast

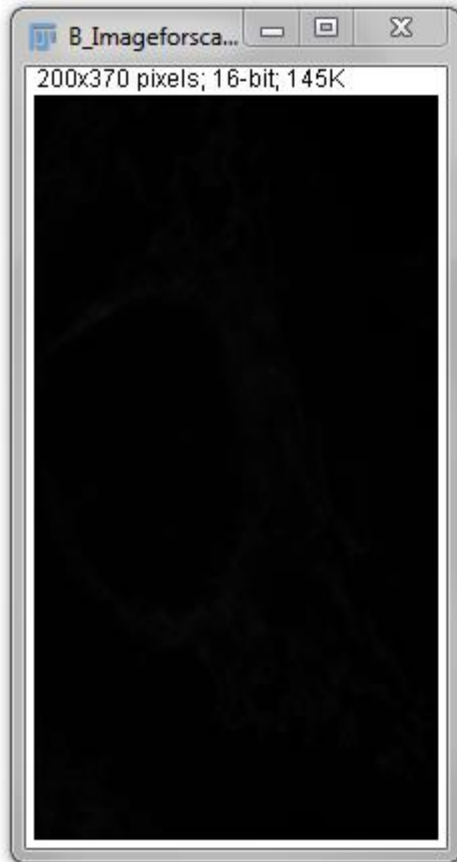
Min to Max

Type in numbers,
can set to all open images

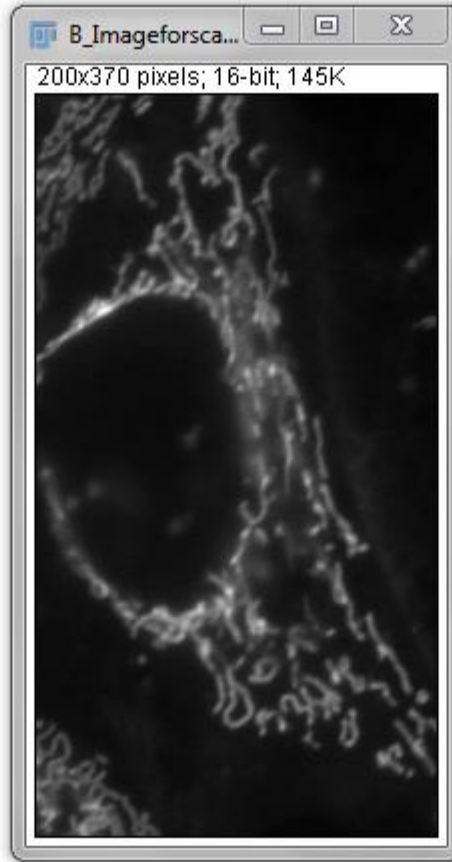
Changes the pixels!

Contrast in image scaling

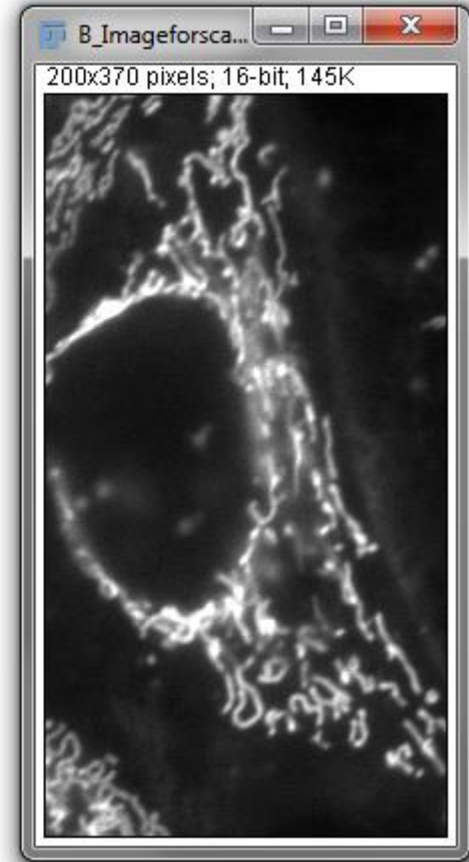
Full range of CCD



Full range of image



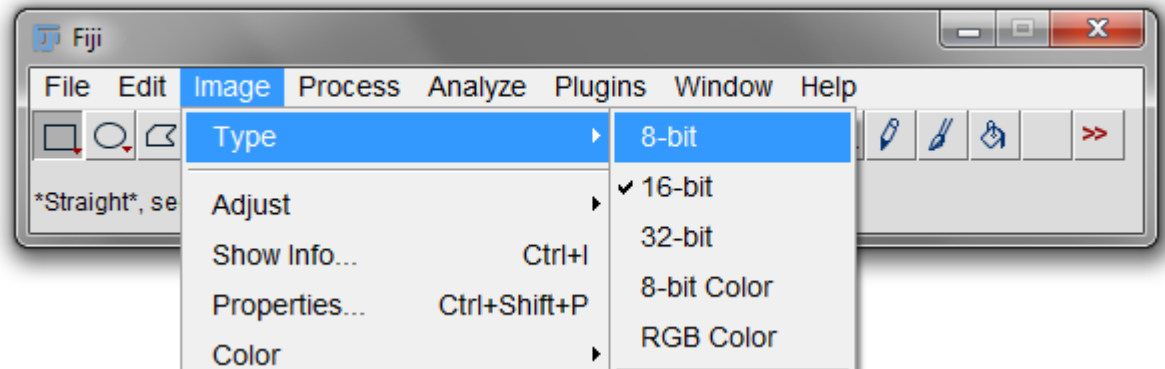
Slight clipping



Why is this useful,
when is this ok,
how much is ok?

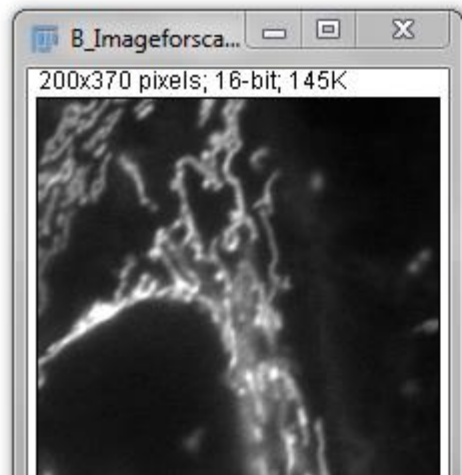
Exporting an 8-bit copy

Ctrl+shift+D
=Duplicate

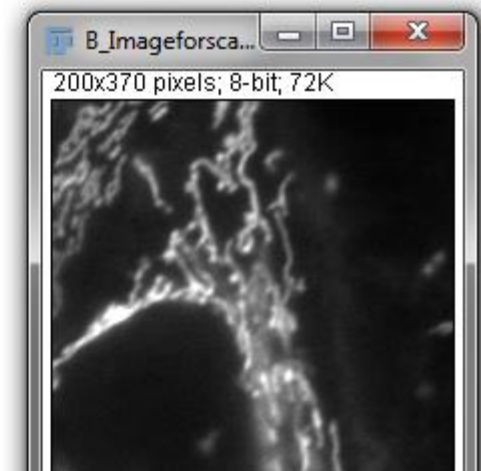


Save your original images -
use for quantification

Easy display/publication



Original pixel intensity
range stretched or
compressed to 8-bit



Which format should I use for export?

Probably makes sense to use the format of the microscope as much as possible, if you do need to export to a standard format . . .

TIFF (aka TIF), and variants

- Lossless
- Suitable bit-depths (8-bit or 24-bit RGB for export)
- Space for metadata (not always populated properly)
- Open
- Pretty much universal

What happens to the numbers?



Scaling has limitations

Scaled to 0 - 4095

100 msec

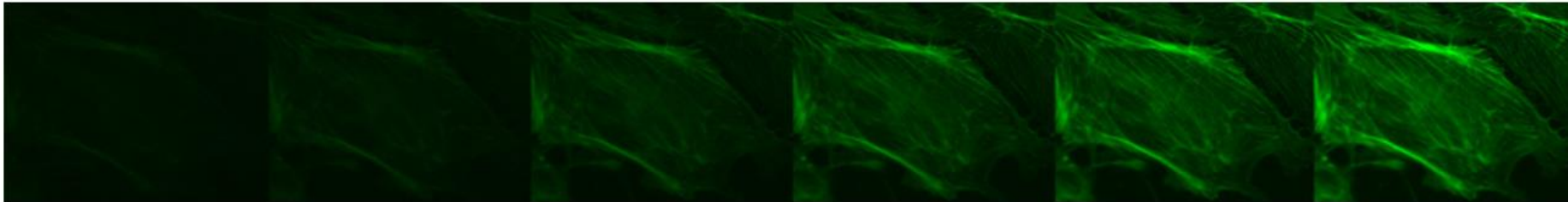
200 msec

400 msec

600 msec

800 msec

1000 msec



100 msec

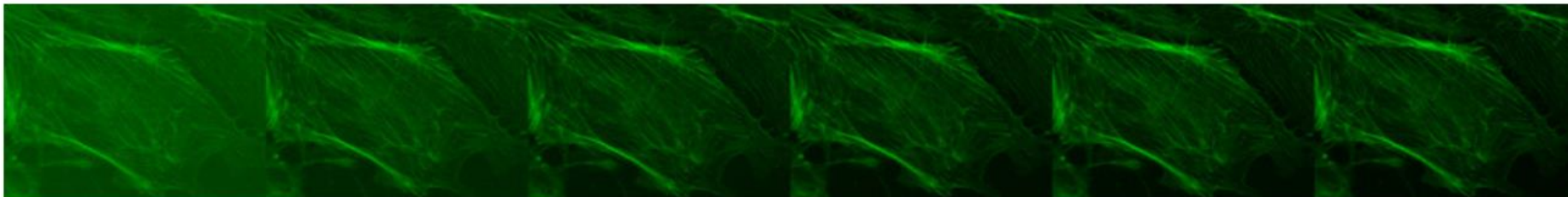
200 msec

400 msec

600 msec

800 msec

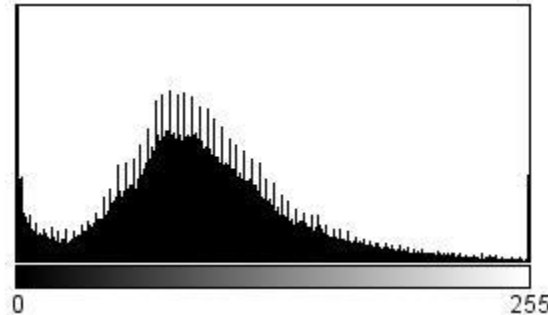
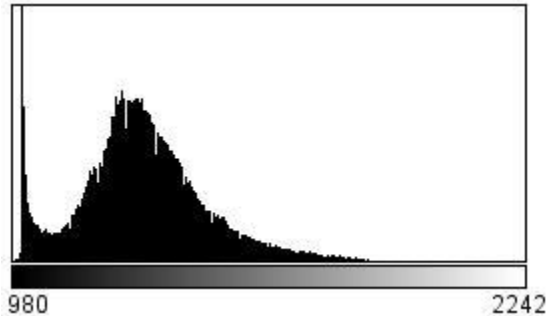
1000 msec



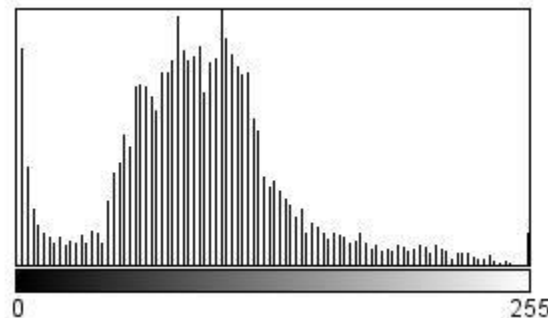
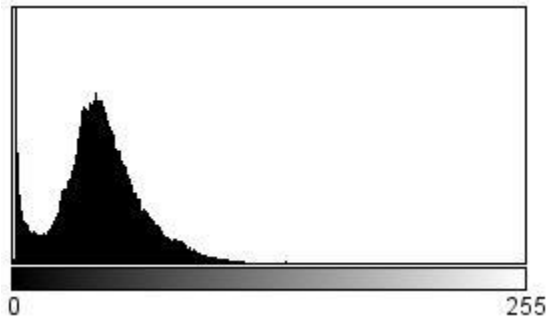
Auto-scaled to min-max of each image

How many grey-values should you have in your raw images?

Imperfections in image scaling



12-bit to 8-bit
compression



8-bit to 8-bit
Stretch

Avoid scaling any images you want to quantify



Exercise: Fair display

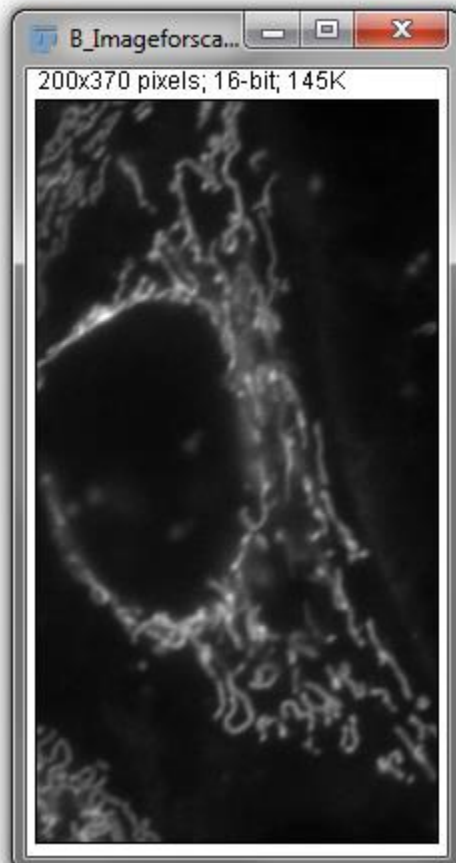
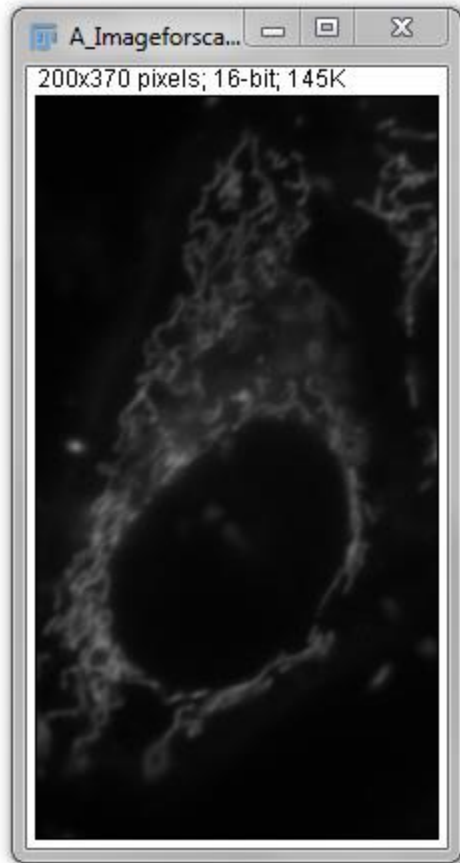
Files:

A_Imageforscaling

B_Imageforscaling

Display these to:

1. Reflect the intensities
 2. Show best structure
- Make 8-bit copies of each pair





Inverting an image: making a negative

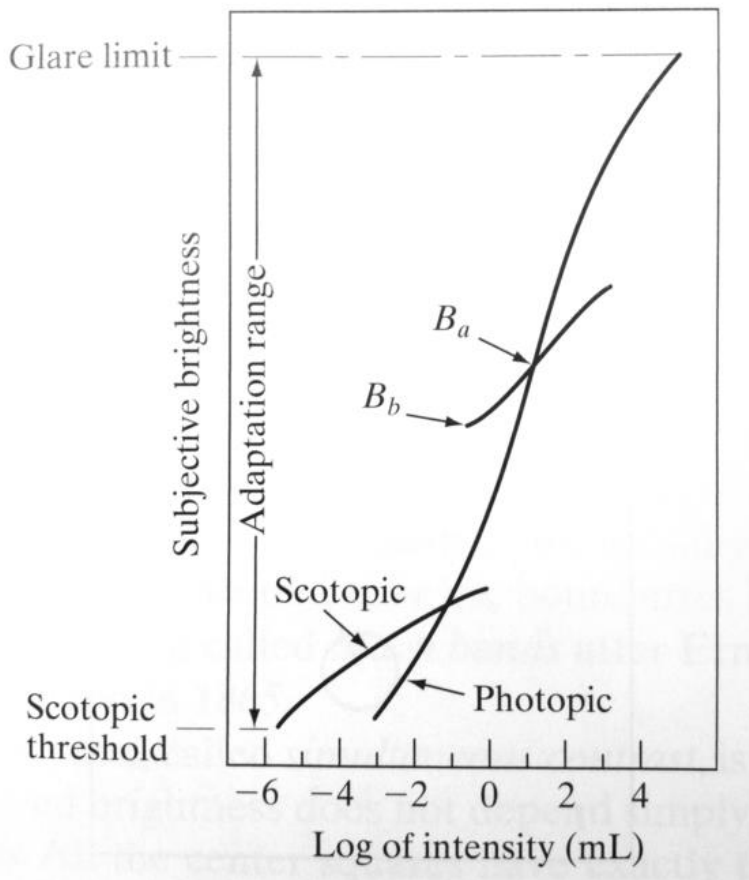
```
Files:  
Invert_this.tif  
A_Imageforscaling
```

- Try inverting an image: Edit/Invert
- Look at the histogram
- Mouse-over the pixels
- Scale it
- Compare to original

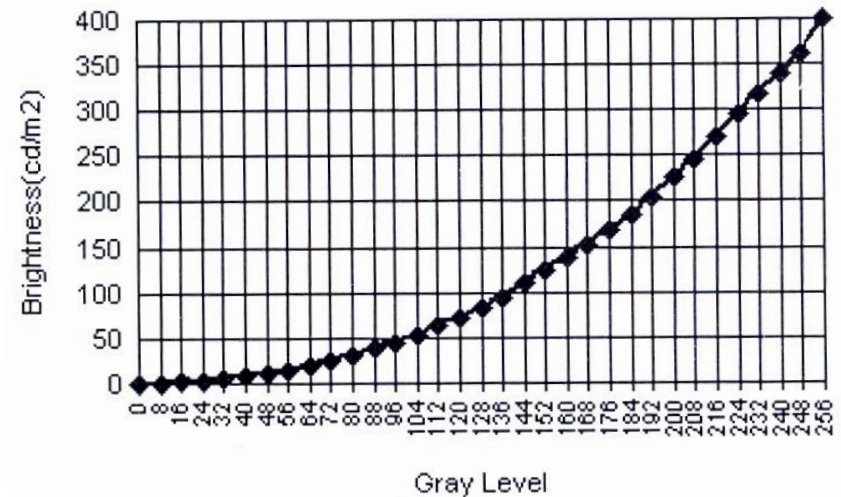
Linearity in intensity display

That sounds a good idea, let's do that

Problem



Solution



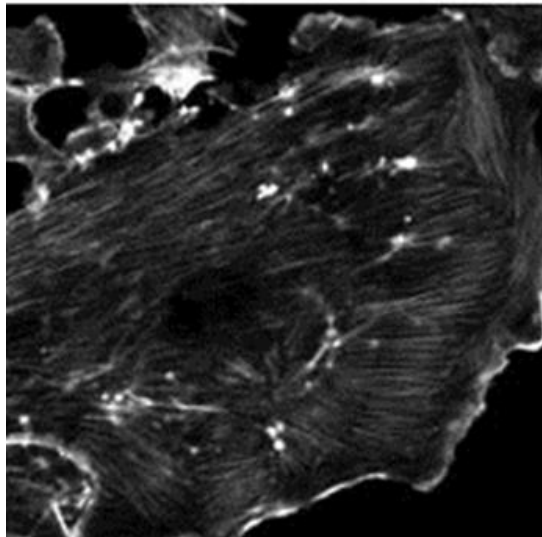


File: gamma

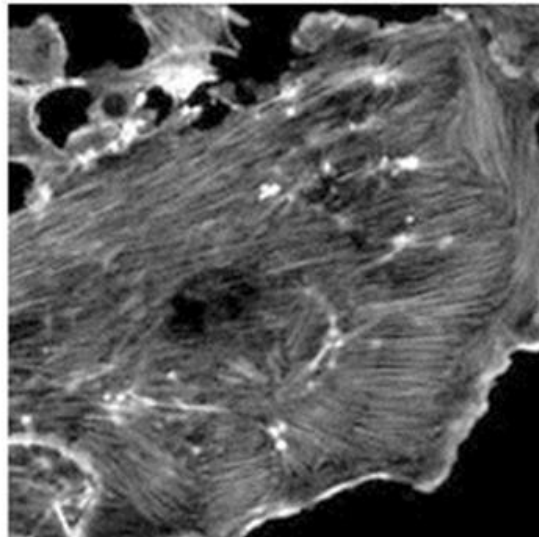
Gamma

output = input ^{γ}

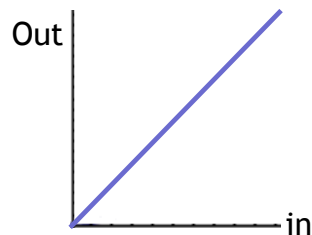
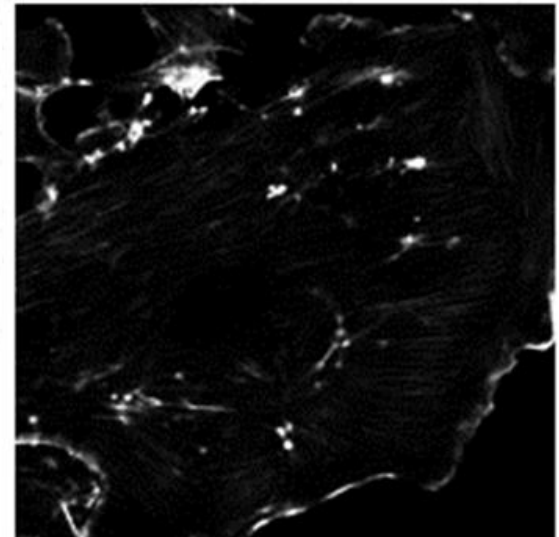
$\gamma = 1$



$\gamma = 0.5$



$\gamma = 2$



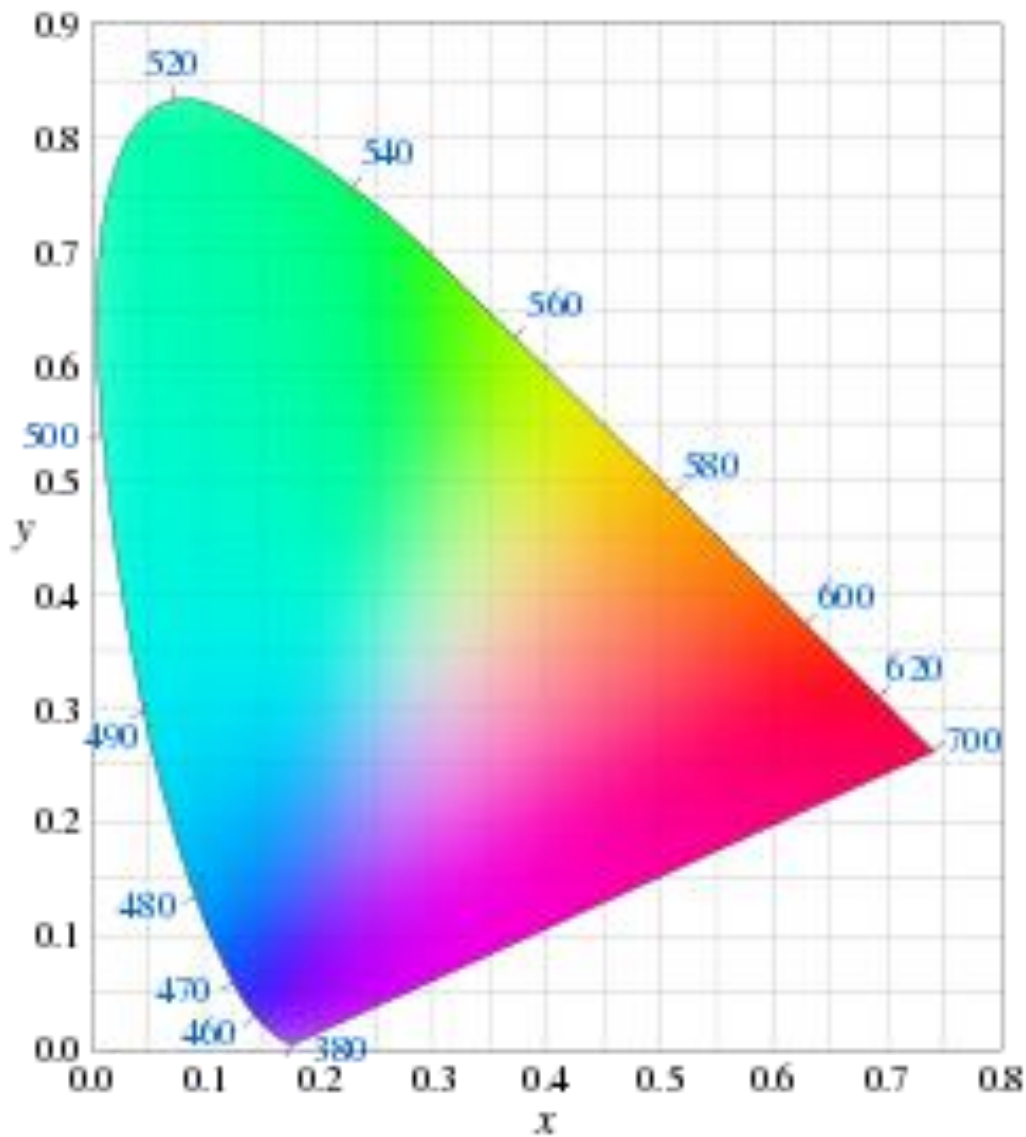
“Linear” display of brightness: $\gamma = 1$

Stick with 1 if you can, certainly state if any images are not linear

You can change your image: </process/math/gamma>

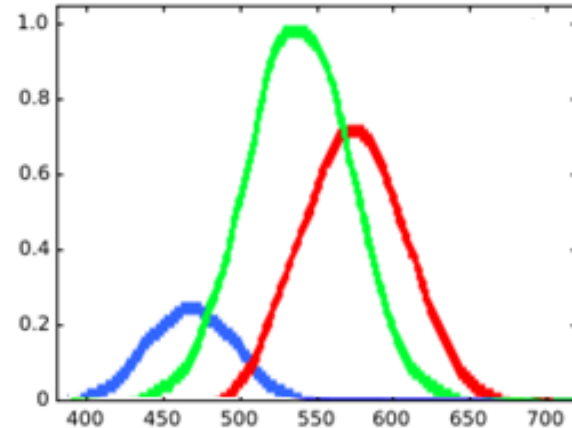
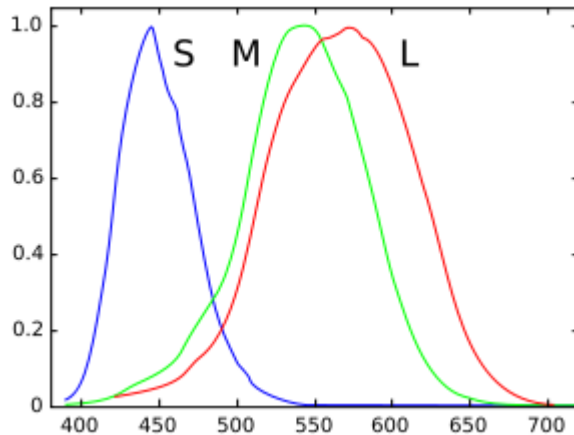
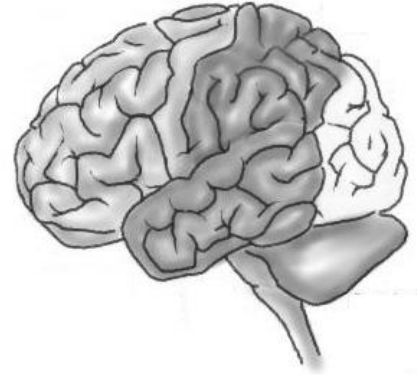
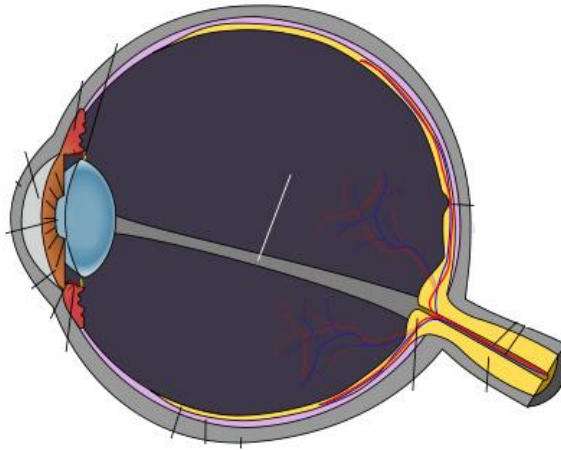
Color

The 10 million colors we can see



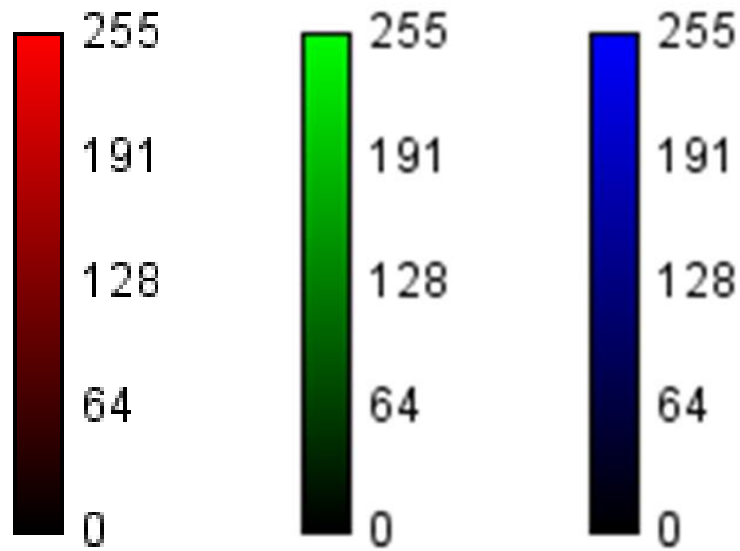
Our eyes

How many
Mpx are
these?



Monitors etc fit in with our eyes - tiny little R, G or B pixels

24-bit RGB



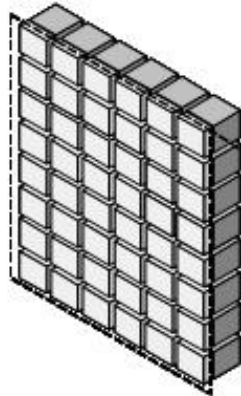
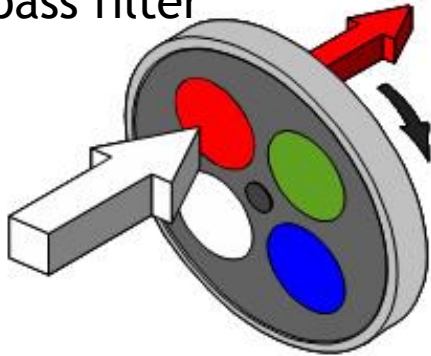
$$256^3 = 16.77 \text{ million}$$

Color in imaging

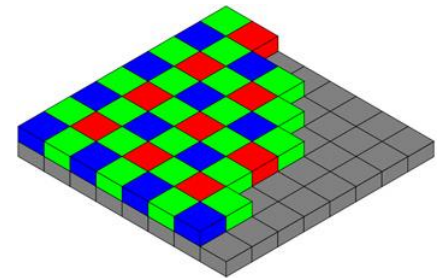
Most of it is actually pseudo-color in fluorescence:

Sensitive monochrome detector

Bandpass filter

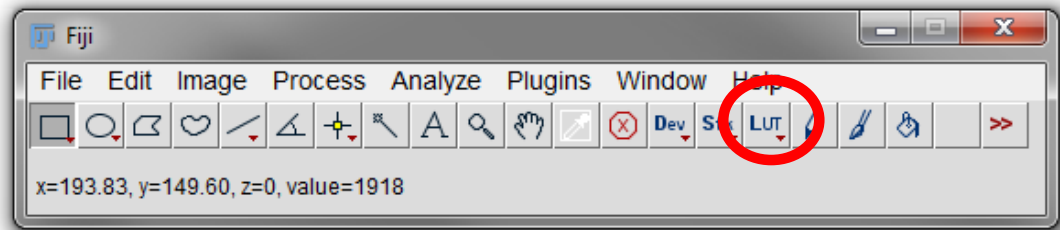


Bayer mask in color camera

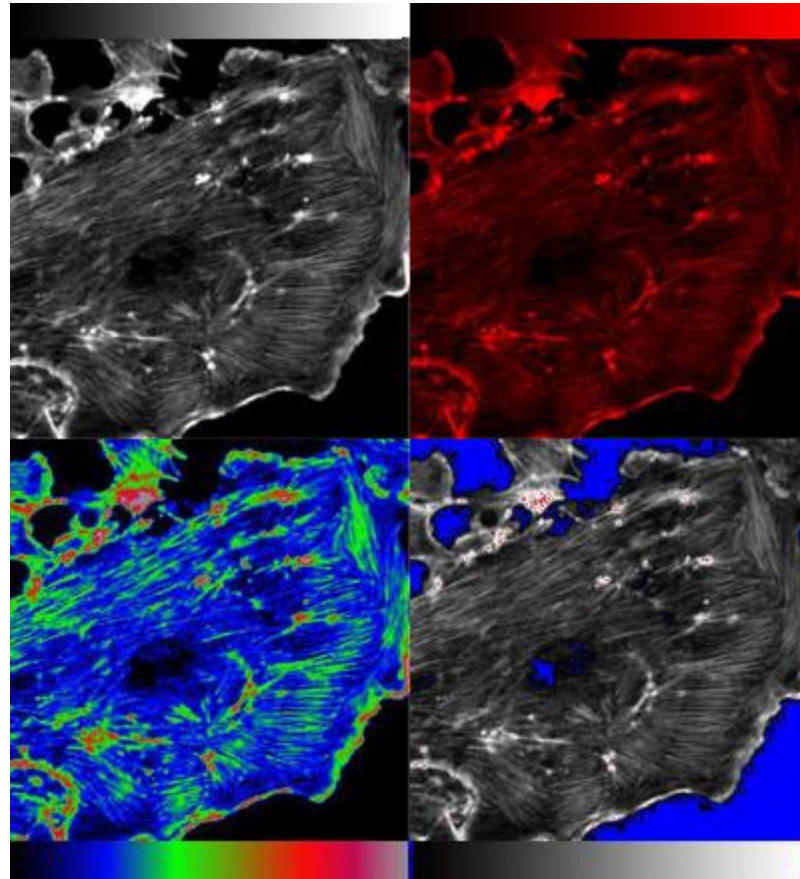


We use color to add context and aid interpretation

Lookup tables



Different ways of displaying the same image . . .



Helpful for distinguishing
similar intensities

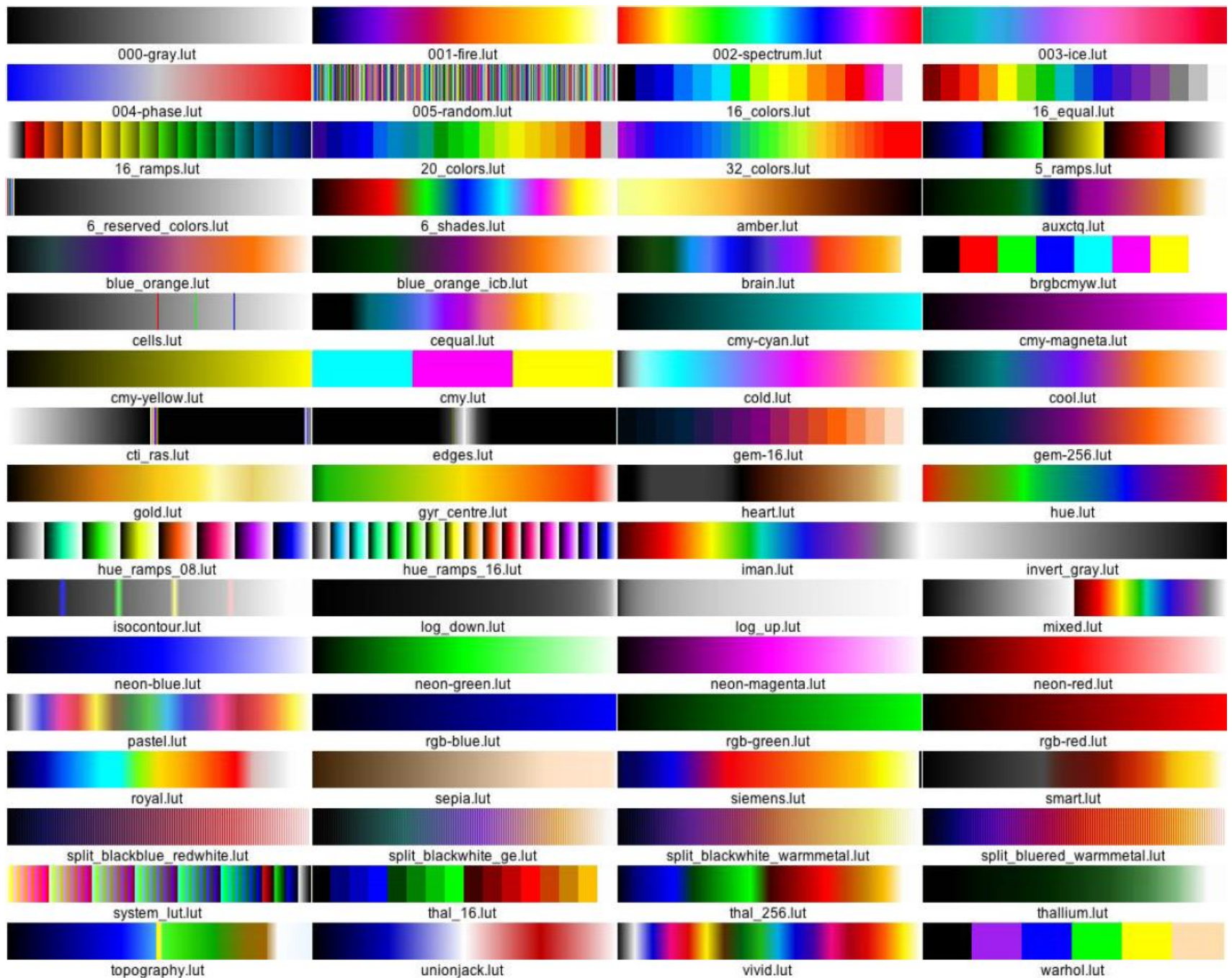
Good for adjusting the
gain and offset on
the confocal



Try some LUTs

File: gamma

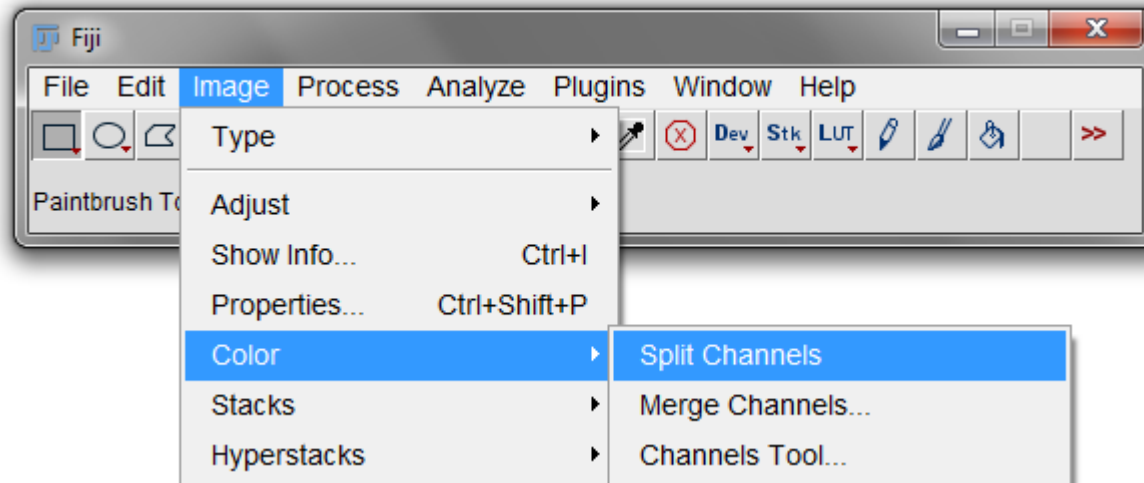
- Try lots of different ones
- Analyze/Tools/Calibration bar
- Inverted LUT - same as image inversion? Mouse over.
- Do you think inverted is a good way of displaying?





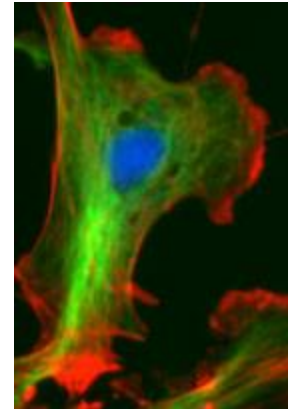
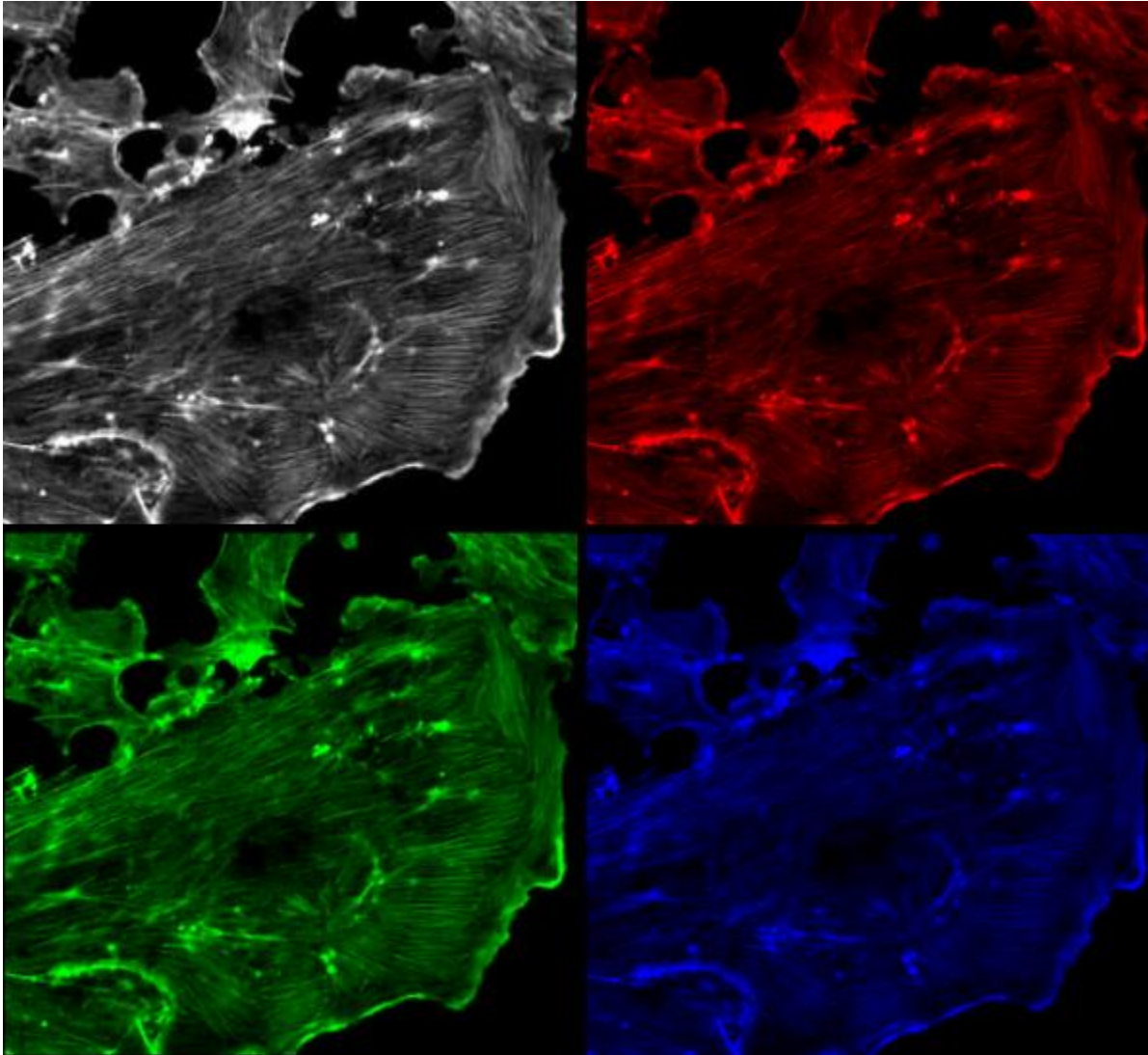
Color display

File:
FluorescentCells

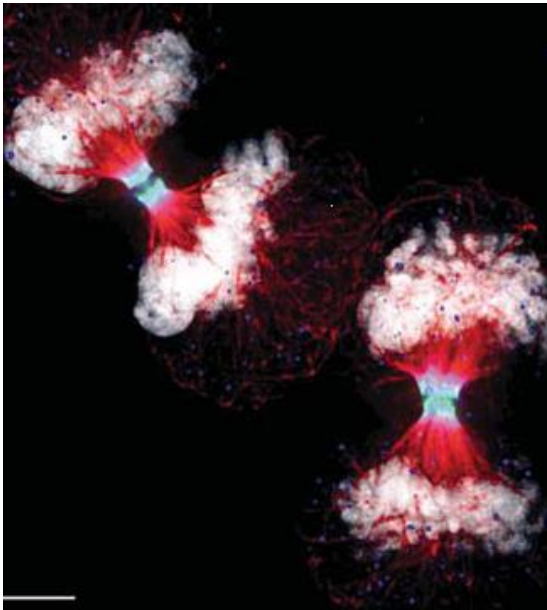


- ✓ RGB and Composite (more channels, more flexible)
- ✓ Channels Tool
- ✓ Split and Merge

Color display isn't always useful



4 or more color images





Exercise: Color display

- ✓ RGB and Composite (more channels, more flexible)
- ✓ Channels Tool
- ✓ Split and Merge

```
Files:  
ImageQuad_A  
ImageQuad_B  
ImageQuad_C  
ImageQuad_DAPI
```

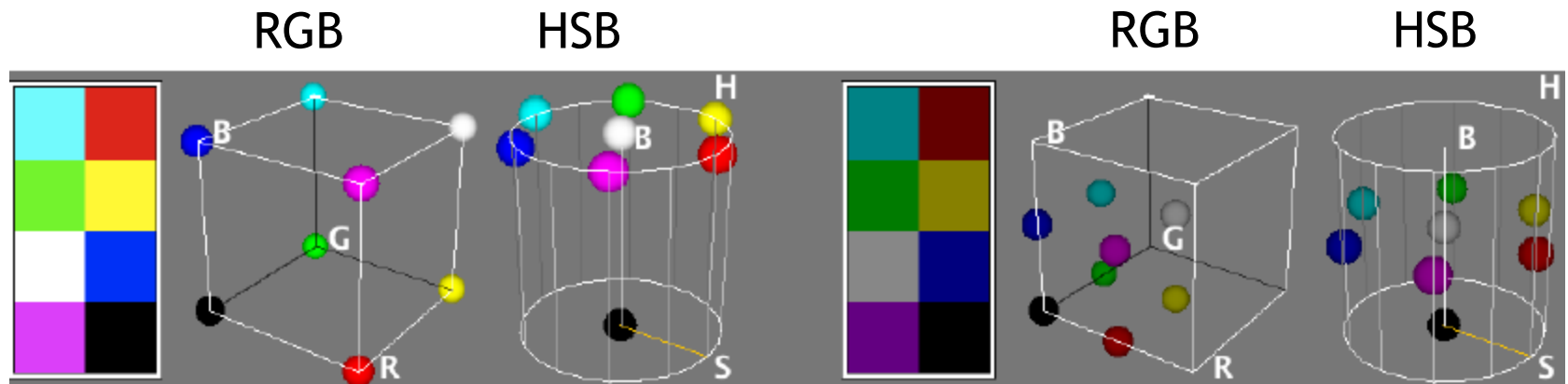
Take these 4 channels of the same cell, make a composite with the channels pseudo-colored and scaled as you think best displays them and export as 24-bit RGB tiff

Should you scale the 4-channels in the same way?
Can you split the final RGB back to your 4 images?



Other color spaces

File:
H&E.tiff



- Image/type/HSB stack or RGB stack
- Image/Adjust/Threshold Colour